



Morphological and phylogenetic evaluation of a new species of *Rhodoveronaea* (Rhamphoriaceae, Rhamphoriales) associated with *Quercus fabrei* (Fagales) in Yunnan Province, China

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Abstract

Background

During an extensive mycological exploration of Fagales-associated mycoflora within the biodiverse landscapes of Yunnan Province, China, we discovered a new saprotrophic fungal species. Based on its unique morphological characteristics and phylogenetic analyses of combined LSU, ITS, SSU, *tef1*-α and *rpb2* sequences, we established

Rhodoveronaea querci as a new member of the genus *Rhodoveronaea*, which was collected from *Quercus fabrei* colonising terrestrial habitats.

New information

Rhodoveronaea querci is currently only known from its asexual form, which is a morphologically distinct hyphomycete fungus. The distinguishing characteristics of *R. querci* are straight or slightly flexuous, septate and unbranched conidiophores, terminally, integrated, apically protuberant, recurved conidiogenous cells with conspicuous conidiogenous loci and ellipsoidal to narrowly-obovoid, mostly 1-septate, slightly constricted at the central septum, guttulate, conidia with a flat basal scar. *Rhodoveronaea querci* and *R. hainanensis* are highly similar in the size and shape of their conidiophores and conidia, but they can be distinguished by other morphological characteristics and by genetic differences. *Rhodoveronaea* species are widely distributed in Europe and Asia and recent studies revealed several new *Rhodoveronaea* species from China.

Keywords

new species, asexual morph, Fagaceae, phylogeny, taxonomy

Introduction

Rhodoveronaea Arzanlou, W. Gams & Crous was introduced by Arzanlou et al. (2007) with *R. varioseptata* Arzanlou, W. Gams & Crous as the type species. The genus showed a close phylogenetic association with the family Annulatascaceae, based on 28S rDNA sequences. Later, Réblová and Štěpánek (2018) introduced the family Rhamphoriaceae to accommodate *Rhodoveronaea* along with three more genera including, *Rhamphoria* Niessl, *Rhamphoriopsis* Réblová & Gardiennet and *Xylolentia* Réblová. Later, the family was placed in its own order Rhamphoriales that was established, based on multigene phylogenetic analysis and divergence time estimates (Hyde et al. 2021).

The asexual morph of *Rhodoveronaea* is characterised by reddish-brown, straight or flexuose conidiophores with inflated basal cells, terminally integrated conidiogenous cells with crowded, slightly conspicuous conidium-bearing denticles, pale brown, ellipsoidal to obovoidal and septate conidia with a protruding base and a marginal basal frill (Arzanlou et al. 2007, Luo et al. 2019, Yuan et al. 2020, Yang et al. 2023, Chen et al. 2024). The sexual morph is distinguished by immersed ascomata with subglobose to conical venter bearing conical neck, filamentous, septate paraphyses longer than asci, unitunicate, cylindrical asci with long-stipitate and fusiform, septate, hyaline ascospores (Réblová 2009, Luo et al. 2019). To date, *Rhodoveronaea varioseptata* is the only known sexual morph within the genus. The asexual morph of *R. varioseptata* was initially identified on *Bertia moriformis* (Tode) De Not. in Germany and the sexual morph was later

documented on *Carpinus betulus* L. in the Czech Republic (now Czechia) (Réblová 2009).

Currently, this genus includes seven species: *Rhodoveronaea aquatica* Z.L. Luo, K.D. Hyde & Hong, *R. everniae* Crous & Boers, *R. hainanensis* Jian Ma, K.D. Hyde & Y.Z. Lu, *R. hyalina* X.M. Chen & Karun., *R. lignicola* X.M. Chen & Tibpromma, *R. nieuwwulvenica* Crous & Osieck and *R. varioseptata*. Amongst these, four species were collected in China, whereas the remaining were described from the Netherlands and the Czechia (Réblová 2009, Crous et al. 2023, Hyde et al. 2023a, Chen et al. 2024). These taxa have been found on different hosts in terrestrial and freshwater habitats (Arzanlou et al. 2007, Réblová 2009, Luo et al. 2019, Crous et al. 2021, Crous et al. 2023, Hyde et al. 2023a, Yang et al. 2023, Chen et al. 2024). Notably, *Rhodoveronaea aquatica* and *R. hainanensis* are specifically known from freshwater habitats in China, while the other species have been collected from terrestrial habitats (Luo et al. 2019, Hyde et al. 2023b).

Despite their occurrence in diverse ecological niches, the diversity and ecological roles of *Rhodoveronaea* species within the order Fagales, particularly in the context of the family Fagaceae, remain largely unexplored. Although the ecological significance of Fagaceae, research on the fungal diversity associated with this family remains limited, particularly regarding saprobic fungi and their ecological interactions (Wang et al. 2011, Li et al. 2012, Jiang et al. 2022, Phukhamsakda et al. 2022, Jayawardena et al. 2023, Yang et al. 2023, Hyde et al. 2024).

During a comprehensive survey of microfungi associated with Fagales in Yunnan Province, China, we encountered a previously undocumented species of *Rhodoveronaea* on decayed wood of *Quercus fabrei* Hance (Thiyagaraja et al. 2024). Based on integrated morphological and multi-locus phylogenetic analyses, this isolate was recognised as a novel taxon. This discovery enriches the taxonomic framework of *Rhodoveronaea* and sheds light on the microfungal diversity within Fagaceae in Yunnan, an area known for its rich biodiversity.

Materials and methods

Sample collection, morphological study and isolation

Dead sticks of *Quercus fabrei* were collected during a survey of Fagales associated microfungi in Kunming City, Yunnan Province, China during the rainy season (October 2021). Kunming, located in a typical subtropical plateau monsoon climate zone, is characterised by mild temperatures throughout the year. The annual average temperature is approximately 15°C and the annual precipitation averages 1000 mm, with most rainfall occurring between May and October. The city's average altitude is around 1,891 m, contributing to its distinct climate features. The vegetation types in this region mainly include evergreen broad-leaved forests, coniferous forests, shrublands and grasslands, as well as artificial forests and farmland, forming a diverse ecological environment that provides favourable conditions for fungal growth. The two specimens

used in this study were collected from the naturally growing Fagaceae forest in Heilongtan Park in the northern area of Kunming City, with loci of 25.143730N, 102.749002E, 25.142479N, 102.753244E, respectively (Fig. 1).

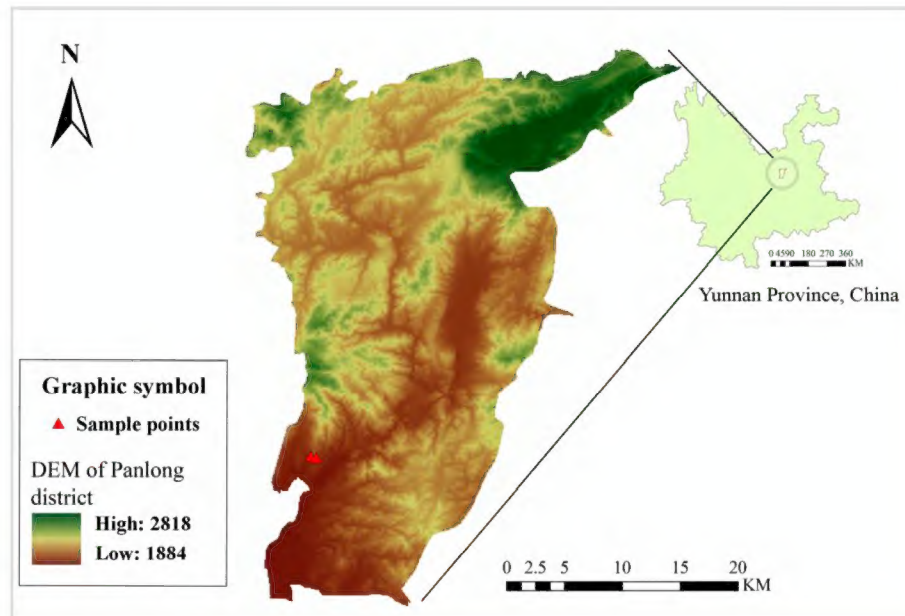


Figure 1. [doi](#)

Map of the northern urban area of Kunming City with altitude information.

Samples were packed in sealed bags and taken to the laboratory for further observation. Materials were examined with a Nikon SMZ 745T stereomicroscope and the macromorphological characters of the hyphomycetes were photographed using a IMG SC2000C camera. Fruiting bodies were picked up with a needle and placed on a slide with distilled water for observation of microscopic characters (Senanayake et al. 2020). The microscopic characteristics were observed using a Nikon ECLIPSE Ni-U microscope (Model Eclipse Ni-UNikon Corporation Tokyo, Japan) and the structures of conidiophores, conidiogenous cells and conidia were photographed by the charge-coupled camera (IMG SC2000C, Jiangsu Province, China). The photo plates were edited with Adobe Photoshop 2019 and measurements were taken using the Tarosoft (R) Image Frame Work programme. A pure culture was obtained by single spore isolation as described by Senanayake et al. (2020). The germinated spores were transferred to a fresh potato dextrose agar (PDA) plate and grew to 2.5 cm in diameter after being incubated at 25–27°C for two months. The characteristics of the culture were frequently checked. Dry-herbarium specimens and living cultures were deposited in the Kunming Institute of Botany Academia Sinica (HKAS) and Kunming Institute of Botany Culture Collection (KUMCC), respectively. Index Fungorum number and Faces of Fungi number were registered as outlined by Index Fungorum (2024) and Jayasiri et al. (2015). The data are also deposited in the Greater Mekong Subregion database (Chaiwan et al. 2021) and Fungalpedia webpage (Hyde et al. 2023b).

DNA extraction, PCR amplification and sequencing

Mycelia were scraped from the growing culture and DNA was extracted using a Trelief™ Plant Genomic DNA extraction kit (TsingKe, Beijing, China) following the manufacturer's

instructions. The large subunit ribosomal rRNA (LSU), internal transcribed spacer (ITS) region, small subunit of the nuclear ribosomal DNA (SSU) and translation elongation factor (*tef1-α*) were amplified with primers LR0R/LR5 (Vilgalys and Hester 1990), ITS1F/ITS4 (White et al. 1990, Gardes and Bruns 1993), NS1/NS4 (Rehner and Samuels 1994) and ef1-983F/ef1-2218R (Rehner and Samuels 1994), respectively. The PCR reactions contained 21 µl of 1×PCR Master Mix, 1 µl of each forward and reverse primer and 2 µl of genomic DNA. The following thermo-cycling parameters were used for ITS, LSU, SSU and *tef1-α*: initially, 95°C for 3 min, followed by 35 cycles of denaturation at 95°C for 15 s, annealing at 54°C for 15 s, elongation at 72°C for 20 s and final extension at 72°C for 5 min. The amplified PCR products were sequenced in Tsingke (Kunming, China). Generated sequences were deposited in GenBank, and accession numbers were obtained.

Sequence alignment and phylogenetic analyses

The newly-generated forward and reverse sequences were assembled using Sequencing Project Management (SeqMan) (Clewley 1995). The combined sequences were subjected to BLASTn search at the National Center for Biotechnology Information (NCBI) to identify closely-related taxa. Related species were downloaded from GenBank following recent publications (Table 1) (Chen et al. 2024). The sequence of individual gene was aligned using MAFFT v. 7 (Kato et al. 2019) and the aligned sequences were trimmed using TrimAl v. 1.2 with default settings (Castresana 2000, Talavera and Castresana 2007). Sequences of all genes were combined using Sequence Matrix v. 1.7.8 (Vaidya et al. 2011). Phylogenetic analyses were performed following Dissanayake et al. (2020). Maximum Likelihood (ML) analysis of the combined multi-gene dataset was performed using RAXML-HP2 on XSEDE v. 8.2.12 in the CIPRES Science Gateway v. 3.3 (<https://www.phylo.org>) with 1000 bootstrap iterations and GTR+I+G substitution model. For Bayesian Inference (BI) analysis, MrModelTest v. 2.2 (Nylander 2004) was used in the Akaike Information Criterion (AIC) to obtain the best-fit evolutionary model of each genetic marker. The posterior probabilities (PP) were performed by Markov Chain Monte Carlo sampling (MCMC) in MrBayes v. 3.1.2. Six simultaneous Markov chains were run for 1,000,000 generations and trees were sampled every 100th generation. The first 25% of trees were discarded in the burn-in phase, while the remaining 75% were used to calculate PP in the majority rule consensus tree (Hua et al. 2006). The generated tree was viewed in FigTree v. 1.4.0 (<http://tree.bio.ed.ac.uk/software/figtree>) and saved for editing in Adobe Illustrator 2020 (Adobe Inc., United States).

Table 1. GenBank accession numbers of the taxa used in the phylogenetic analyses in this study.							
Taxa	Strain/ Voucher	GenBank Accession Numbers					References
		LSU	ITS	SSU	<i>tef1-α</i>	<i>rpb2</i>	
<i>Myrmecridium schulzeri</i>	CBS 100.54	EU041826	EU041769	NA	NA	NA	Arzanlou et al. (2007)

Taxa	Strain/ Voucher	GenBank Accession Numbers					References
		LSU	ITS	SSU	<i>tef1-α</i>	<i>rpb2</i>	
<i>Neomyrmecridium sorbicola</i>	CBS 143433	MH107948	MH107901	NA	NA	NA	Crous et al. (2018)
<i>Rhamphoria aquimicrospora</i>	GZCC 20-0515 ^T	OP377911	OP377812	OP377996	OP472992	OP473085	Yang et al. (2023)
<i>R. hyalospora</i>	MFLU 19-2849 ^T	MN846342	MN846344	NA	NA	NA	Lin et al. (2023)
<i>R. muriformis</i>	CBS 131269 ^T	MG600396	NA	MG600404	NA	MG600399	Réblová and Štěpánek (2018)
<i>R. muriformis</i>	CBS 127683	MG600395	MG600389	MG600403	NA	MG600400	Réblová and Štěpánek (2018)
<i>R. pyriformis</i>	CBS 139024 ^T	MG600397	MG600392	MG600405	NA	MG600401	Réblová et al. (2016)
<i>R. pyriformis</i>	CBS 139033	KT991665	KT991677	MG600406	NA	KT991656	Réblová et al. (2016)
<i>R. sympodialis</i>	HKAS 105172 ^T	MT079191	MT079187	NA	NA	NA	Hyde et al. (2020)
<i>Rhodoveronaea aquatica</i>	MFLUCC 18-1339 ^T	MK849785	MK828641	MK828310	MN194046	NA	Luo et al. (2019)
<i>R. aquatica</i>	GZCC 20-0447	OP377947	OP377862	OP378027	OP473041	OP473107	Yang et al. (2023)
<i>R. everniae</i>	CBS 148309 ^T	OK663776	OK664737	NA	NA	OK651172	Crous et al. (2021)
<i>R. hainanensis</i>	GZAAS 22-2020 ^T	OP748932	OP748935	NA	NA	NA	Hyde et al. (2023a)
<i>R. hyalina</i>	GZCC 23-0622 ^T	PP102207	PP102206	PP102214	PP259403	PP259399	Chen et al. (2024)
<i>R. hyalina</i>	GZCC 23-0623	PP102208	PP102211	PP102215	PP259404	PP259400	Chen et al. (2024)
<i>R. lignicola</i>	GZCC 23-0624 ^T	PP102209	PP102212	PP102216	PP259405	PP259401	Chen et al. (2024)
<i>R. lignicola</i>	GZCC 23-0625	PP102210	PP102213	PP102217	PP259406	PP259402	Chen et al. (2024)
<i>R. nieuwwulvenica</i>	CBS 149447 ^T	OQ629048	OQ628466	NA	OQ627955	OQ627935	Crous et al. (2023)
<i>R. querci</i>	HKAS 145561^T	PV097144	PQ932528	PV097142	PV164602	NA	This study
<i>R. querci</i>	HKAS 145562	PV097145	PQ932529	PV097143	PV164603	NA	This study
<i>R. varioseptata</i>	CBS 431.88 ^T	EU041870	EU041813	NA	NA	NA	Arzanlou et al. (2007)

Taxa	Strain/ Voucher	GenBank Accession Numbers					References
		LSU	ITS	SSU	<i>tef1-α</i>	<i>rpb2</i>	
<i>R. varioseptata</i>	CBS 123472	FJ617559	MG600393	MG600408	NA	JX066701	Arzanlou et al. (2007)
<i>R. varioseptata</i>	CBS 123473	FJ617560	KT991676	JX066710	NA	JX066700	Arzanlou et al. (2007)
<i>Xylolentia aseptata</i>	GZCC 20-0424 ^T	OP377944	OP377859	OP378024	OP473038	OP473104	Yang et al. (2023)
<i>X. reniformis</i>	MFLU 19-0210 ^T	MK547648	MK547646	NA	NA	NA	Yuan et al. (2020)

Taxon treatment

Rhodoveronaea querci Y. Y. Yang & Q. Zhao, sp. nov.

- MycoBank [859526](#)

Material

Holotype:

- a. kingdom: Fungi; phylum: Ascomycota; class: Sordariomycetes; order: Rhamphoriales; family: Rhamphoriaceae; genus: *Rhodoveronaea*; country: China; stateProvince: Yunnan; county: Kunming; year: 2021; month: October; day: 6; habitat: terrestrial; catalogNumber: HKAS 145561; occurrenceRemarks: on dead sticks of *Quercus fabrei*; recordNumber: YYY79; recordedBy: Y. Y. Yang; identifiedBy: Y. Y. Yang; institutionCode: Herbarium of Kunming Institute of Botany, Academia Sinica (KUN-HKAS); occurrenceID: E2970614-75ED-5C09-9C9D-0208B4B658C1

Description

Saprobic on dead sticks of *Quercus fabrei*. **Sexual morph**: Undetermined. **Asexual morph**: Hyphomycetous. *Colonies* on natural substrate superficial, effuse, hairy, scattered or aggregated, with yellowish-brown conidial masses at the apex. *Mycelium* mostly immersed, partly superficial, composed of filamentous, septate, branched, brown to pale brown hyphae. *Conidiophores* 85–129 μm × 3–4.5 μm (\bar{x} = 101 × 3.5 μm, n = 15), macronematous, mononematous, erect, simple, cylindrical, straight or slightly flexuous, solitary, septate, unbranched, dark-brown, pale brown towards fertile apex, slightly rough-walled, often percurrently proliferating from cut ends. *Conidiogenous cells* polyblastic, integrated, indeterminate, terminal, sympodial, pale brown, cylindrical below, apically protuberant, recurved with conspicuous conidiogenous loci. *Conidia* 8–14 μm × 3–5.5 μm (\bar{x} = 11 × 4 μm, n = 30), acropleurogenous, ellipsoidal to narrowly obovoid, apically rounded, with a flat basal scar, pale brown, 1–2-septate, mostly 1-septate, slightly constricted at the central septum, guttulate, smooth-walled. *Conidial secession* schizolytic.

Culture characteristics: Conidia germinating on PDA within 24 h. Colonies slow growing on PDA media, reaching 2.5 cm after 2 months at 18°C, offwhite, irregularly-shaped with obvious concentric rings, raised, pleated, with irregular edges, with sparse mycelia; reverse greige, with white mycelial ring.

Diagnosis

Similar to *R. hainanensis* in conidial size and morphology, but differing in the thickness of the conidiophore wall, as well as the septation and guttulation of the conidia.

Etymology

The species epithet "*querci*" refers to the host genus.

Notes

In the phylogenetic analyses, our species formed a sister clade to *R. aquatica*, *R. hainanensis* and *R. nieuwwulvenica* (Fig. 2). Pairwise comparisons revealed that *R. querci* differs from *R. aquatica* by 23/506 bp (4.5%) in the ITS sequences, 6/695 bp (0.8%) in the LSU sequences, 0/712 bp (0%) in the SSU sequences and 36/776 bp (4.6%) in the *tef1-α* sequences. Furthermore, *R. querci* differs from *R. hainanensis* by 18/460 bp (3.9%) in the ITS sequences and 12/721 bp (1.7%) in the LSU sequences. Additionally, *R. querci* differs from *R. nieuwwulvenica* by 23/490 bp (4.7%) in the ITS sequences, 9/721 bp (1.2%) in the LSU sequences and 38/744 bp (5.1%) in the *tef1-α* sequences. Morphologically (Table 2), *Rhodoveronaea querci* resembles *R. aquatica*, *R. hainanensis* and *R. nieuwwulvenica* by having cylindrical, septate, straight or flexuous, red-brown conidiophores, as well as ellipsoid to obovoid, pale brown conidia (Luo et al. 2019, Hyde et al. 2023a, Crous et al. 2023). However, *R. querci* can be distinguished from *R. aquatica* by its shorter conidiophores (85–129 μm × 3–4.5 μm vs. 182–310 × 9–13 μm) and smaller conidia (8–14 μm × 3–5.5 μm vs. 23–27 × 9–11 μm) (Luo et al. 2019). In contrast to *R. hainanensis*, *R. querci* possesses integrated conidiogenous cells and conidia with guttulate cells and fewer septa (1–2-septate vs. 1–3-septate) (Hyde et al. 2023a). Furthermore, the difference between *R. querci* and *R. nieuwwulvenica* is that *R. nieuwwulvenica* has flexuous, shorter conidiophores (20–60 × 4–5 μm vs. 85–129 μm × 3–4.5 μm) and more septate conidia (3-septate vs. 1–2-septate) (Crous et al. 2023). We therefore introduce this strain as a new species following the guidelines of Chethana et al. (2021).

Analysis

Phylogenetic trees were constructed using 25 isolates, including two newly-generated sequences (Table 1). The concatenated dataset consisted of 4614 characters including gaps (LSU: 1-870 bp, ITS: 871-1403 bp, SSU 1404-2708 bp, *tef1-α*: 2709-3605 bp and

rpb2: 3606-4614 bp). *Myrmecridium schulzeri* (CBS 100.54) and *Neomyrmecridium sorbicola* (CBS 143433) were used as outgroup taxa following Chen et al. (2024). The final optimised likelihood value of the ML tree with the highest score was -16176.373461. The alignment had 1073 distinct alignment patterns, with 35.50% of undetermined characters and gaps. The parameters for the GTR+I+G model of combined LSU, ITS, SSU, *tef1- α* and *rpb2* were as follows: estimated base frequencies A = 0.251898, C = 0.251105, G = 0.274700, T = 0.222296; substitution rates AC = 1.481862, AG = 2.986319, AT = 1.330144, CG = 1.336388, CT = 9.252404, GT = 1.000000; and gamma distribution shape parameter α = 0.636081. Phylogenetic analysis showed that *Rhodoveronaea querci* clustered with the other three *Rhodoveronaea* species and formed a basal lineage with 62% ML support (Fig. 3).



Figure 2. [doi](#)

Rhodoveronaea querci (HKAS 145561, holotype). **a** Host substrate; **b, c** Colonies on the substrate; **d, e** Conidiophores and conidiogenous cells bearing conidia; **f, g** Conidiogenous cells bearing conidia. Note the percurrent proliferation of conidiophores from cut-ends in e-g; **h-m** Conidia; **n** Germinated conidium; **o, p** Upper and lower view of culture on PDA media after incubation for 60 days. Scale bars: **d-e, g, n** = 30 μ m, **f** = 20 μ m, **h-m** = 10 μ m.

Table 2.

Synopsis table of prominent morphological and culture characteristics for *Fitzroyomyces* species.

Species Name	Conidiophores	Conidiogenous cells	Conidia	Culture	References
<i>Rhodoveronaea aquatica</i>	182–310 × 9–13 µm, cylindrical, straight or flexuose, thick-walled, septate, red-brown, paler at apex.	Polyblastic, terminally integrated, sympodial, smooth.	23–27 × 9–11 µm, acropleurogenous, ellipsoid to obovoid, apically rounded, with a flat basal scar, 1–3-septate, pale brown.	No	Luo et al. (2019)
<i>R. everniae</i>	25–100 × 3–4 µm, subcylindrical, straight to geniculate-sinuous, branched or not, thick-walled, warty, 1–6-septate, red-brown.	15–40 × 3–4 µm, integrated, terminal and intercalary, smooth to warty, forming a rachis of minute pimple-like subdenticulate loci.	(6.5–)9–11(– 12) × (3–)4(–4.5) µm, ellipsoid to obovoid, with a protruding hilum and minute marginal frill, 0–3-septate, pale brown.	Colonies erumpent, spreading, with moderate aerial mycelium and smooth, lobate margin.	Crous et al. (2021)
<i>R. hainanensis</i>	58–172 µm × 5.5–10.5 µm, cylindrical, straight or flexuose, thick-walled, septate, red-brown, paler at apex.	Polyblastic, terminally integrated, sympodial, sometimes branched, smooth.	10–15 µm × 4–5.5 µm, acropleurogenous, ellipsoid to obovoid, apically rounded, with a flat basal scar, 1–3-septate, pale brown.	Colonies circular, flat, red brown at the entire margin, white in the centre, reverse-side red-brown.	Hyde et al. (2023a)
<i>R. hyalina</i>	95–165 × 4.5–8 µm, cylindrical, erect or geniculate-sinuous, smooth-walled, septate, unbranched, red-brown, paler towards the apex.	Polyblastic, integrated, terminal, determinate, sympodial.	14–18 × 4.5–5.5 µm, acropleurogenous, ellipsoidal to narrowly obovoid, apically rounded with a flat basal scar, aseptate, hyaline to subhyaline, guttulate.	Colonies circular, with an umbonate surface, edge regular, obvious rings, rings dark brown, the rest of the grey-brown, reverse black, with inconspicuous circles.	Chen et al. (2024)
<i>R. lignicola</i>	75–125 × 3.5–7.5 µm, cylindrical, straight or slightly flexuous, slightly rough-walled, septate, unbranched, dark brown, paler towards the apex.	Polyblastic, integrated, terminal, determinate, sympodial.	9–13.5 × 4–6 µm, acropleurogenous, ellipsoidal to narrowly obovoid, conical at apex with a flat basal scar, 1–3-septate when mature, hyaline to pale yellowish, guttulate.	Colonies circular white to greyish-white, with an umbonate and irregular edge, reverse light yellow, with inconspicuous circles	Chen et al. (2024)

Species Name	Conidiophores	Conidiogenous cells	Conidia	Culture	References
<i>R. nieuwwulvenica</i>	20–60 × 4–5 µm, subcylindrical, flexuous, thick-walled, 2–3-septate, red-brown.	20–50 × 4–5 µm, integrated, terminal, finely verruculose, forming a rachis with subdenticulate loci.	(8–)11–13(–14) × (3–)4(–4.5) µm, solitary, subcylindrical to narrowly fusoid-ellipsoid, apex subobtuse, base bluntly rounded, 3-septate, medium brown, guttulate.	Colonies erumpent, folded, with sparse aerial mycelium and smooth, lobate margin.	Crous et al. (2023)
<i>R. querci</i>	85–129 µm × 3–4.5 µm, cylindrical, straight or slightly flexuous, slightly rough-walled, septate, unbranched, dark-brown, pale brown towards fertile apex, often percurrently proliferating from cut ends.	Polyblastic, integrated, indeterminate, terminal, sympodial, apically protuberant, recurved with conspicuous conidiogenous loci.	8–14 µm × 3–5.5 µm, acropleurogenous, ellipsoidal to narrowly obovoid, apically rounded, with a flat basal scar, 1–2-septate, pale brown, guttulate.	Colonies off-white with obvious concentric rings, raised, pleated, with irregular edges, with sparse mycelia, reverse greige, with white mycelial ring.	This study
<i>R. varioseptata</i>	80–93 × 4–5 µm, cylindrical, straight or flexuous, brown to red-brown.	Polyblastic, integrated, sympodial, bearing up to 15 sympodially produced denticles.	14–16 × 5–6.5 µm, ellipsoidal to obovoidal, apically rounded, with a flat basal scar and marginal frill, 0–3(–4)-septate, pale brown.	Colonies surface olivaceous-grey to olivaceous-pale brown with a pale grey zone at the margin, aerial mycelium dense, whitish to pale grey, margin entire, reverse olivaceous-black.	Réblová (2009)

Discussion

In this study, we introduce a new saprobic species of *Rhodoveronaea* associated with Fagales, based on phylogenetic studies of combined LSU, ITS, SSU, *tef1-α* and *rpb2* datasets, as well as morphological details. Previous studies by Arzanlou et al. (2007) employed LSU and ITS sequences to classify *Rhodoveronaea*. More recently, multi-gene datasets, such as LSU-ITS-*tef1-α-rpb2* and LSU-ITS-SSU-*tef1-α-rpb2*, have enhanced the phylogenetic resolution of the genus (Crous et al. 2023, Hyde et al. 2023a). However, some species still lack gene fragments, as in *R. everniae* and *R. hainanensis*, which only have data on ITS and LSU sequences (Crous et al. 2021, Hyde et al. 2023a). In future studies, it will be necessary to collect and sequence more gene regions of *Rhodoveronaea* samples.

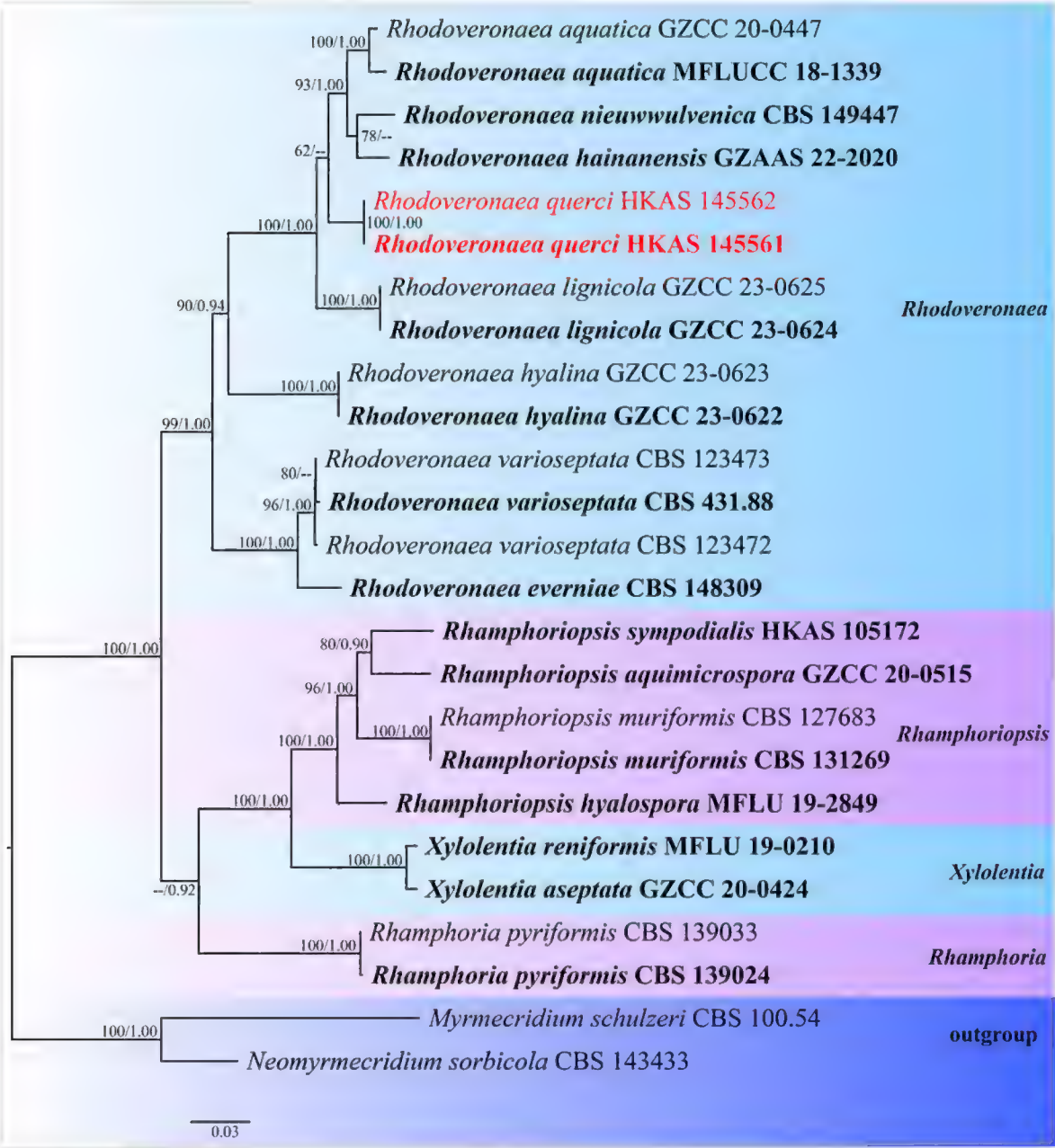


Figure 3. [doi](#)
RAxML tree, based on combined LSU, ITS, SSU, *tef1-α* and *rpb2* dataset analyses. Bootstrap support values for Maximum Likelihood (ML, first value) equal to or greater than 60% and Bayesian posterior probabilities (BPP, second value) equal to or greater than 0.90 are given at the nodes. Ex-type strains are in bold; the newly-generated isolates are in red.

In our phylogenetic trees, *Rhodoveronaea querci* formed a distinct clade sister to *R. aquatica*, *R. nieuwwulvenica* and *R. hainanensis* with low bootstrap support (62 ML/0.68 PP). This relationship was consistent across repeated phylogenetic analyses, with significant differences observed in their nucleotide comparisons (Table 3). Morphologically, *Rhodoveronaea querci* is similar to *R. hainanensis* and *R. aquatica* in having unbranched, red-brown conidiophores with septate, ellipsoidal to narrowly obovoid, pale brown and guttulate conidia, but differed from the latter by having flexuous conidiogenous cells with protuberant, slightly recurved apices and conidia with more septa (1–2-septate vs. 1–3-septate) (Luo et al. 2019). *Rhodoveronaea querci* shares similarities with *R. nieuwwulvenica* in having terminal, integrated conidiogenous cells and comparably similar-sized guttulate conidia (8–14 × 3–5.5 μm vs. 8–14 × 3–4.5 μm). However, it can be distinguished by its larger conidiophores (85–129 × 3–4.5 μm vs. 20–60 × 4–5 μm) and fewer septa in conidia (1–2-septate vs. 3-septate) (Crous et al. 2023).

Table 3.
Base differences in *Rhodoveronaea querci* compared to other species. “NA”: no data available in GenBank.

Species	Strain	ITS	LSU	SSU	<i>tef1-α</i>
<i>R. aquatica</i>	MFLUCC 18-1339	23/506 bp (4.5%)	6/695 bp (0.8%)	0/712 bp (0%)	36/776 bp (4.6%)
<i>R. hainanensis</i>	GZAAS 22-2020	18/460 bp (3.9%)	12/721 bp (1.7%)	NA	NA
<i>R. nieuwwulvenica</i>	CBS 149447	23/490 bp (4.7%)	9/721 bp (1.2%)	NA	38/744 bp (5.1%)

Rhodoveronaea species have been reported on different hosts, such as *Bertia moriformis* (fungal substrate), *Evermia prunastri* (L.) Ach. (Parmeliaceae), *Carpinus betulus* (Betulaceae) and Bambusoideae (Arzanlou et al. 2007, Réblová 2009, Crous et al. 2021, Crous et al. 2023). In this study, *Rhodoveronaea* was reported for the first time on *Quercus fabrei*. The species of *Rhodoveronaea* are widely distributed in Asia and Europe and most species are collected in the temperate region, with only a few from tropical and subtropical regions (Arzanlou et al. 2007, Réblová 2009, Luo et al. 2019, Crous et al. 2021, Crous et al. 2023, Hyde et al. 2023a). Due to the limited ecological and distributional data available, it remains uncertain whether host-specificity and climatic conditions influence the growth of *Rhodoveronaea* species. Collecting *Rhodoveronaea* species poses significant challenges due to their minute, hyphomycetous morphology and inconspicuous growth habits, which often render them overlooked in field studies. Currently, five species have been reported in China and two species have been found on Fagales. *Rhodoveronaea varioseptata* (CBS 123472) and *R. querci* were isolated as saprobes from *Carpinus betulus* (Betulaceae, Fagales) and *Quercus fabrei* (Fagaceae, Fagales), respectively (Réblová 2009, Luo et al. 2019, Hyde et al. 2023a, Chen et al. 2024). The differences between *R. varioseptata* and *R. querci* are the conidiogenous cells of *R. varioseptata* have up to 15 sympodially produced denticles producing holoblastic conidia holoblastically and the conidia are slightly larger than the latter (Réblová 2009). This study expands our understanding of the diversity and host associations of *Rhodoveronaea*, highlighting the potential for discovering new fungal species on Fagales plants, particularly in understudied regions and habitats. Further investigations may provide insights into the ecological roles and evolutionary history of these fungi.

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References

- Arzanlou M, Groenewald JZ, Gams W, Braun U, Shin H-D, Crous PW (2007) Phylogenetic and morphotaxonomic revision of *Ramichloridium* and allied genera. *Studies in Mycology* 58: 57-93. <https://doi.org/10.3114/sim.2007.58.03>
- Castresana J (2000) Selection of conserved blocks from multiple alignments for their use in phylogenetic analysis. *Molecular Biology and Evolution* 17 (4): 540-552. <https://doi.org/10.1093/oxfordjournals.molbev.a026334>
- Chaiwan N, Gomdola D, Wang S, Monkai J, Tibpromma S, Doilom M, Wanasinghe DN, Mortimer PE, Lumyong S, Hyde KD (2021) <https://gmsmicrofungi.org>: an online database providing updated information of microfungi in the Greater Mekong Subregion. *Mycosphere* 12 (1): 1513-1526. <https://doi.org/10.5943/mycosphere/12/1/19>
- Chen XM, Tang X, Du TY, Lu YZ, Suwannarach N, Xu RF, Yang EF, Elgorban AM, Tibpromma S, Karunarathna SC (2024) Two new species of *Rhodoveronea* (Rhamphoriales, Ascomycota) from terrestrial habitats in China. *Phytotaxa* 638 (1): 49-60. <https://doi.org/10.11646/phytotaxa.638.1.4>
- Chethana KT, Manawasinghe IS, Hurdeal VG, Bhunjun CS, Appadoo MA, Gentekaki E, Raspé O, Promputtha I, Hyde KD (2021) What are fungal species and how to delineate them? *Fungal Diversity* 109 (1): 1-25. <https://doi.org/10.1007/s13225-021-00483-9>
- Clewley JP (1995) Macintosh sequence analysis software: DNASTar's LaserGene. *Molecular Biotechnology* 3 (3): 221-224. <https://doi.org/10.1007/bf02789332>
- Crous PW, Schumacher RK, Wingfield MJ, Akulov A, Denman S, Roux J, Braun U, Burgess TI, Carnegie AJ, Váczy KZ, Guatimosim E, Schwartsburd PB, Barreto RW, Hernández-Restrepo M, Lombard L, Groenewald JZ (2018) New and Interesting Fungi. 1. *Fungal Systematics and Evolution* 1 (1): 169-215. <https://doi.org/10.3114/fuse.2018.01.08>
- Crous PW, Osieck ER, Jurjevi Ž, Boers J, Van Iperen AL, Starink-Willemse M, Dima B, Balashov S, Bulgakov TS, Johnston PR, Morozova OV, Pinruan U, Sommai S, Alvarado P, Decock CA, Lebel T, McMullan-Fisher S, Moreno G, Shivas RG, Zhao L, Abdollahzadeh J, Abrinbana M, Ageev DV, Akhmetova G, Alexandrova AV, Altés A, Amaral AG, Angelini C, Antonín V, Arenas F, Asselman P, Badali F, Baghela A, Bañares A, Barreto RW, Baseia IG, Bellanger J-M, Berraf-Tebbal A, Biketova AY, Bukharova NV, Burgess TI, Cabero J, Câmara MP, Cano-Lira JF, Ceryngier P, Chávez R, Cowan DA, de Lima AF, Oliveira RL, Denman S, Dang QN, Dovana F, Duarte IG, Eichmeier A, Erhard A, Esteve-Raventós F, Fellin A, Ferisin G, Ferreira RJ, Ferrer A, Finy P, Gaya E, Geering AD, Gil-Durán C, Glässnerová K, Glushakova AM, Gramaje D, Guard FE, Guarnizo AL, Haelewaters D, Halling RE, Hill R, Hirooka Y, Hubka V, Iliushin VA, Ivanova DD, Ivanushkina NE, Jangsantear P, Justo A, Kachalkin AV, Kato S, Khamsuntorn P, Kirtsideli IY, Knapp DG, Kochkina GA, Koukol O, Kovács GM, Kruse J, Kumar TK, Kušan I,

- Læssøe T, Larsson E, Lebeuf R, Levicán G, Loizides M, Marinho P, Luangsa-ard JJ, Lukina EG, Magaña-Dueñas V, Maggs-Kölling G, Malysheva EF, Malysheva VF, Martín B, Martín MP, Matočec N, McTaggart AR, Mehrabi-Koushki M, Mešić A, Miller AN, Mironova P, Moreau P-, Morte A, Müller K, Nagy LG, Nanu S, Navarro-Ródenas A, Nel WJ, Nguyen TH, Nóbrega TF, Noordeloos ME, Olariaga I, Overton BE, Ozerskaya SM, Palani P, Pancorbo F, Papp V, Pawłowska J, Pham TQ, Phosri C, Popov ES, Portugal A, Pošta A, Reschke K, Reul M, Ricci GM, Rodríguez A, Romanowski J, Ruchikachorn N, Saar I, Safi A, Sakolrak B, Salzmann F, Sandoval-Denis M, Sangwichein E, Sanhueza L, Sato T, Sastoque A, Senn-Irlet B, Shibata A, Siepe K, Somrithipol S, Spetik M, Sridhar P, Stchigel AM, Stuskova K, Suwannasai N, Tan YP, Thangavel R, Tiago I, Tiwari S, Tkalčec Z, Tomashevskaya MA, Tonegawa C, Tran HX, Tran NT, Trovão J, Trubitsyn VE, Van Wyk J, Vieira WA, Vila J, Visagie CM, Vizzini A, Volobuev SV, Vu DT, Wangsawat N, Yaguchi T, Ercole E, Ferreira BW, de Souza AP, Vieira BS, Groenewald JZ (2021) Fungal Planet description sheets: 1284–1382. *Persoonia - Molecular Phylogeny and Evolution of Fungi* 47 (1): 178-374. <https://doi.org/10.3767/persoonia.2021.47.06>
- Crous PW, Osieck ER, Shivas RG, Tan YP, Bishop-Hurley SL, Esteve-Raventós F, Larsson E, Luangsa-ard JJ, Pancorbo F, Balashov S, Baseia IG, Boekhout T, Chandranayaka S, Cowan DA, Cruz RH, Czachura P, De la Peña-Lastra S, Dovana F, Drury B, Fell J, Flakus A, Fotedar R, Jurjević Ž, Kolečka A, Mack J, Maggs-Kölling G, Mahadevakumar S, Mateos A, Mongkolsamrit S, Noisripoom W, Plaza M, Overy DP, Pitek M, Sandoval-Denis M, Vauras J, Wingfield MJ, Abell SE, Ahmadpour A, Akulov A, Alavi F, Alavi Z, Altés A, Alvarado P, Anand G, Ashtekar N, Assyov B, Banc-Prandi G, Barbosa KD, Barreto GG, Bellanger JM, Bezerra JL, Bhat DJ, Bilański P, Bose T, Bozok F, Chaves J, Costa-Rezende DH, Danteswari C, Darmostuk V, Delgado G, Denman S, Eichmeier A, Etayo J, Eyssartier G, Faulwetter S, Ganga KG, Ghosta Y, Goh J, Góis JS, Gramaje D, Granit L, Groenewald M, Gulden G, Gusmão LF, Hammerbacher A, Heidarian Z, Hywel-Jones N, Jankowiak R, Kaliyaperumal M, Kaygusuz O, Kezo K, Khonsanit A, Kumar S, Kuo CH, Laessøe T, Latha KP, Loizides M, Luo SM, Maciá-Vicente JG, Manimohan P, Marbach PA, Marinho P, Marney TS, Marques G, Martín MP, Miller AN, Mondello F, Moreno G, Mufeeda KT, Mun HY, Nau T, Nkomo T, Okrańska A, Oliveira JP, Oliveira RL, Ortiz DA, Pawłowska J, Pérez-De-Gregorio MÀ, Podile AR, Portugal A, Privitera N, Rajeshkumar KC, Rauf I, Rian B, Rigueiro-Rodríguez A, Rivas-Torres GF, Rodriguez-Flakus P, Romero-Gordillo M, Saar I, Saba M, Santos CD, Sarma PV, Siquier JL, Sleiman S, Spetik M, Sridhar KR, Stryjak-Bogacka M, Szczepańska K, Taşkın H, Tennakoon DS, Thanakitpipattana D, Trovão J., Türkeul A, van Iperen AL, van 't Hof P, Vasquez G, Visagie CM, Wingfield BD, Wong PT, Yang WX, Yarar M, Yarden O, Yilmaz N, Zhang N, Zhu YN, Groenewald JZ (2023) Fungal Planet description sheets: 1478–1549. *Persoonia - Molecular Phylogeny and Evolution of Fungi* 50 (1): 158-310. <https://doi.org/10.3767/persoonia.2023.50.05>
 - Dissanayake AJ, Bhunjun CS, Maharachchikumbura SS, Liu JK (2020) Applied aspects of methods to infer phylogenetic relationships amongst fungi. *Mycosphere* 11 (1): 2652-2676. <https://doi.org/10.5943/mycosphere/11/1/18>
 - Gardes M, Bruns TD (1993) ITS primers with enhanced specificity for basidiomycetes - application to the identification of mycorrhizae and rusts. *Molecular Ecology* 2 (2): 113-118. <https://doi.org/10.1111/j.1365-294x.1993.tb00005.x>

- Hua Z, Lv Q, Ye W, Wong CK, Cai G, Gu D, Ji Y, Zhao C, Wang J, Yang BB, Zhang Y (2006) MiRNA-directed regulation of VEGF and Other Angiogenic Factors under Hypoxia. *PLOS One* 1 (1). <https://doi.org/10.1371/journal.pone.0000116>
- Hyde KD, Jeewon R, Chen YJ, Bhunjun CS, Calabon MS, Jiang HB, Lin CG, Norphanphoun C, Sysouphanthong P, Pem D, Tibpromma S (2020) The numbers of fungi: is the descriptive curve flattening? *Fungal Divers* 103: 219-271. <https://doi.org/10.1007/s13225-020-00458-2>
- Hyde KD, Bao DF, Hongsanan S, Chethana KWT, Yang J, Suwannarach N (2021) Evolution of freshwater Diaporthomycetidae (Sordariomycetes) provides evidence for five new orders and six new families. *Fungal Diversity* 107 (1): 71-105. <https://doi.org/10.1007/s13225-021-00469-7>
- Hyde KD, Norphanphoun C, Ma J, Yang HD, Zhang JY, Du TY, Gao Y, Gomes de Farias AR, He SC, He YK, Li CJ, Li JY, Liu XF, Lu L, Su HL, Tang X, Tian XG, Wang SY, Wei DP, Xu RF, Xu RJ, Yang YY, Zhang F, Zhang Q, Bahkali AH, Boonmee S, Chethana KW, Jayawardena RS, Lu YZ, Karunarathna SC, Tibpromma S, Wang Y, Zhao Q (2023a) *Mycosphere* notes 387–412 – novel species of fungal taxa from around the world. *Mycosphere* 14 (1): 663-744. <https://doi.org/10.5943/mycosphere/14/1/8>
- Hyde KD, Amuhénage TB, Apurillo CC, Asghari R, Aumentado HD, Bahkali AH, Bera I, Bhunjun CS, Calabon MS, Chandrasiri S, Chethana KW, Doilom M, Dong W, Fallahi M, Kapov S, Khyaju S, Le L, Li CJ, Li QR, Li YX, Lin CG, Linn MM, Liu JK, Liu NG, Luangharn T, Madagammana AD, Manawasinghe IS, Marasinghe DS, McKenzie EH, Meakala N, Meng QF, Mukhopadhyay S, Norphanphoun C, Pem D, Phukhamsakda C, Sarma V, Selcuk F, Senanayake IC, Shah S, Shu YX, Silva HV, Su HL, Tavakol M, Thakshila SA, Thiyagaraja V, Thongklang N, Tian Q, Tibpromma S, Tun ZL, Ulukapi M, Wang Y, Wannasawang N, Wijayawardene NN, Wimalasena SD, Xiao Y, Xiong YR, Yasanthika WA, Li Q, Dai DQ (2023b) *Fungalpedia*, an illustrated compendium of the fungi and fungus-like taxa. *Mycosphere* 14 (1): 1835-1959. <https://doi.org/10.5943/mycosphere/14/1/22>
- Hyde KD, Wijesinghe SN, Afshari N, Aumentado HD, Bhunjun CS, Boonmee S, Camporesi E, Chethana KW, Doilom M, Dong W, Du TY, Farias AR, Gao Y, Jayawardena RS, Karimi O, Karunarathna SC, Kularathnage ND, Lestari AS, Li CJ, Li YX, Liao CF, Liu XF, Lu L, Lu YZ, Luo ZL, Ma J, Mamarabadi M, Manawasinghe IS, Mapook A, Mi LX, Niranjana M, Senanayake IC, Shen HW, Su HL, Tibpromma S, Xu RJ, Yan JY, Yang YH, Yang YY, Yu FQ, Kang JC, Zhang JY (2024) *Mycosphere* Notes 469–520. *Mycosphere* 15 (1): 1294-1454. <https://doi.org/10.5943/mycosphere/15/1/11>
- Jayasiri SC, Hyde KD, Ariyawansa HA, Bhat J, Buyck B, Cai L, Dai YC, Abd-Elsalam KA, Ertz D, Hidayat I, Jeewon R, Jones EBG, Bahkali AH, Karunarathna SC, Liu JK, Luangsa-ard JJ, Lumbsch HT, Maharachchikumbura SSN, McKenzie EHC, Moncalvo JM, Ghobad-Nejhad M, Nilsson H, Pang KL, Pereira OL, Phillips AJL, Raspé O, Rollins AW, Romero AI, Etayo J, Selçuk F, Stephenson SL, Suetrong S, Taylor JE, Tsui CKM, Vizzini A, Abdel-Wahab MA, Wen TC, Boonmee S, Dai DQ, Daranagama DA, Dissanayake AJ, Ekanayaka AH, Fryar SC, Hongsanan S, Jayawardena RS, Li WJ, Perera RH, Phookamsak R, de Silva NI, Thambugala KM, Tian Q, Wijayawardene NN, Zhao RL, Zhao Q, Kang JC, Promputtha I (2015) The Faces of Fungi database: fungal names linked with morphology, phylogeny and human impacts. *Fungal Diversity* 74 (1): 3-18. <https://doi.org/10.1007/s13225-015-0351-8>

- Jayawardena RS, Hyde KD, Wang S, Sun YR, Suwannarach N, Sysouphanthong P, Abdel-Wahab MA, Abdel-Aziz FA, Abeywickrama PD, Abreu VP, Armand A, Aptroot A, Bao DF, Begerow D, Bellanger JM, Bezerra JDP, Bundhun D, Calabon MS, Cao T, Cantillo T, Carvalho JLVR, Chaiwan N, Chen CC, Courtecuisse R, Cui BK, Damm U, Denchev CM, Denchev TT, Deng CY, Devadatha B, de Silva NI, dos Santos LA, Dubey NK, Dumez S, Fernandez HS, Firmino AL, Gafforov Y, Gajanayake AJ, Gomdola D, Gunaseelan S, He SC, Htet ZH, Kaliyaperumal M, Kemler M, Kezo K, Kularathnaga ND, Leonardi M, Li JP, Liao C, Liu S, Loizides M, Luangharn T, Ma J, Madrid H, Mahadevakumar S, Maharachchikumbura SSN, Manamgoda DS, Martín MP, Mekala N, Moreau P, Mu YH, Pahoua P, Pem D, Pereira OL, Phonrob W, Phukhamsakda C, Raza M, Ren GC, Rinaldi AC, Rossi W, Samarakoon BC, Samarakoon MC, Sarma VV, Senanayake IC, Singh A, Souza MF, Souza-Motta CM, Spielmann AA, Su W, Tang X, Tian XG, Thambugala KM, Thongklang N, Tennakoon DS, Wannathes N, Wei DP, Welti S, Wijesinghe SN, Yang H, Yang Y, Yuan HS, Zhang H, Zhang J, Balasuriya A, Bhunjun CS, Bulgakov TS, Cai L, Camporesi E, Chomnunti P, Deepika YS, Doilom M, Duan WJ, Han SL, Huanraluek N, Jones EBG, Lakshmidhevi N, Li Y, Lumyong S, Luo Z, Khuna S, Kumla J, Manawasinghe IS, Mapook A, Punyaboon W, Tibpromma S, Lu YZ, Yan JY, Wang Y (2023) Fungal diversity notes 1512–1610: taxonomic and phylogenetic contributions on genera and species of fungal taxa. *Fungal Diversity* 117 (1): 1-272. <https://doi.org/10.1007/s13225-022-00513-0>
- Jiang N, Voglmayr H, Xue H, Piao CG, Li Y (2022) Morphology and phylogeny of *Pestalotiopsis* (Sporocadaceae, Amphisphaeriales) from Fagaceae leaves in China. *Microbiology Spectrum* 10 (6). <https://doi.org/10.1128/spectrum.03272-22>
- Katoh K, Rozewicki J, Yamada KD (2019) MAFFT online service: multiple sequence alignment, interactive sequence choice and visualization. *Briefings in Bioinformatics* 20 (4): 1160-1166. <https://doi.org/10.1093/bib/bbx108>
- Li HY, Shen M, Zhou ZP, Li T, Wei YI, Lin Lb (2012) Diversity and cold adaptation of endophytic fungi from five dominant plant species collected from the Baima Snow Mountain, Southwest China. *Fungal Diversity* 54 (1): 79-86. <https://doi.org/10.1007/s13225-012-0153-1>
- Lin CG, Liu JK, Chukeatirote E, Hyde KD (2023) *Rhamphoriopsis hyalospora* sp. nov. associated with decaying wood from China. *Phytotaxa* 598 (3): 245-253. <https://doi.org/10.11646/phytotaxa.598.3.6>
- Luo ZL, Hyde KD, Liu JK, Maharachchikumbura SSN, Jeewon R, Bao DF, Bhat DJ, Lin CG, Li WL, Yang J, Liu NG, Lu YZ, Jayawardena RS, Li JF, Su HY (2019) Freshwater Sordariomycetes. *Fungal Diversity* 99 (1): 451-660. <https://doi.org/10.1007/s13225-019-00438-1>
- Nylander JA (2004) MrModeltest v2. Progr Distrib by author. Available online: <https://github.com/nylander/MrModeltest2> (accessed 13 July 2022).
- Phukhamsakda C, Nilsson RH, Bhunjun CS, de Farias AR, Sun YR, Wijesinghe SN, Raza M, Bao DF, Lu L, Tibpromma S, Dong W, Tennakoon DS, Tian XG, Xiong YR, Karunarathna SC, Cai L, Luo ZL, Wang Y, Manawasinghe IS, Camporesi E, Kirk PM, Promputtha I, Kuo CH, Su HY, Doilom M, Li Y, Fu YP, Hyde KD (2022) The numbers of fungi: contributions from traditional taxonomic studies and challenges of metabarcoding. *Fungal Diversity* 114 (1): 327-386. <https://doi.org/10.1007/s13225-022-00502-3>
- Réblová M (2009) Teleomorph of *Rhodoveronaea* (Sordariomycetidae) discovered and re-evolution of *Pleurophragmium*. *Fungal Diversity* 36: 129-139.

- Réblová M, Fournier J, Štěpánek V (2016) Two new lineages of aquatic ascomycetes: *Atractospora* gen. nov. and *Rubellisphaeria* gen. et sp. nov., and a sexual morph of *Myrmecridium montsecurinum* sp. nov. *Mycological Progress* 15 (3). <https://doi.org/10.1007/s11557-016-1166-z>
- Réblová M, Štěpánek V (2018) Introducing the Rhamphoriaceae, fam. nov. (Sordariomycetes), two new genera, and new life histories for taxa with *Phaeoisaria*- and *Idriella*-like anamorphs. *Mycologia* 110 (4): 750-770. <https://doi.org/10.1080/00275514.2018.1475164>
- Rehner SA, Samuels GJ (1994) Taxonomy and phylogeny of *Gliocladium* analysed from nuclear large subunit ribosomal DNA sequences. *Mycological Research* 98 (6): 625-634. [https://doi.org/10.1016/s0953-7562\(09\)80409-7](https://doi.org/10.1016/s0953-7562(09)80409-7)
- Senanayake IC, Rathnayake AR, Marasinghe DS, Calabon MS, Gentekaki E, Lee HB (2020) Morphological approaches in studying fungi: collection, examination, isolation, sporulation and preservation. *Mycosphere* 11 (1): 2678-2754. <https://doi.org/10.5943/mycosphere/11/1/20>
- Talavera G, Castresana J (2007) Improvement of phylogenies after removing divergent and ambiguously aligned blocks from protein sequence alignments. *Systematic Biology* 56 (4): 564-577. <https://doi.org/10.1080/10635150701472164>
- Thiyagaraja V, He SC, Luo L, Meng Q, Yang H, Yang YY (2024) Research Profiles – I: The research of Vinodhini Thiyagaraja and her team at Kunming Institute of Botany, CAS, P.R. China. *Current Research in Environmental & Applied Mycology* 14 (1): 49-62. <https://doi.org/10.5943/cream/14/1/2>
- Vaidya G, Lohman DJ, Meier R (2011) SequenceMatrix: concatenation software for the fast assembly of multi-gene datasets with character set and codon information. *Cladistics* 27 (2): 171-180. <https://doi.org/10.1111/j.1096-0031.2010.00329.x>
- Vilgalys R, Hester M (1990) Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. *Journal of Bacteriology* 172 (8): 4238-4246. <https://doi.org/10.1128/jb.172.8.4238-4246.1990>
- Wang Q, Gao C, Guo LD (2011) Ectomycorrhizae associated with *Castanopsis fargesii* (Fagaceae) in a subtropical forest, China. *Mycological Progress* 10: 323-32. <https://doi.org/10.1007/s11557-010-0705-2>
- White TJ, Bruns T, Lee S, Taylor J (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. *PCR protocols: a guide to methods and applications* 18 (1): 315-322. <https://doi.org/10.1016/b978-0-12-372180-8.50042-1>
- Yang J, Liu LL, Jones EBG, Hyde KD, Liu ZY, Bao DF, Liu NG, Li WL, Shen HW, Yu XD, Liu JK (2023) Freshwater fungi from karst landscapes in China and Thailand. *Fungal Diversity* 119 (1): 1-212. <https://doi.org/10.1007/s13225-023-00514-7>
- Yang YY, Wei DP, Zhao Q, Gomes de Farias AR, Hyde KD (2023) *Neoostropa castaneae* gen. et. sp. nov. (Stictidaceae, Ostropales) and a new report of *Fitzroyomyces cyperacearum* from China. *Current Research in Environmental & Applied Mycology* 13 (1): 123-135. <https://doi.org/10.5943/cream/13/1/9>
- Yuan HS, Lu X, Dai YC, Hyde KD, Kan YH, Kušan I, He SH, Liu NG, Sarma VV, Zhao CL, Cui BK, Yousaf N, Sun G, Liu SY, Wu F, Lin CG, Dayarathne MC, Gibertoni TB, Conceição LB, Garibay-Orijel R, Villegas-Ríos M, Salas-Lizana R, Wei TZ, Qiu JZ, Yu ZF, Phookamsak R, Zeng M, Paloi S, Bao DF, Abeywickrama PD, Wei DP, Yang J, Manawasinghe IS, Harishchandra D, Brahmanage RS, de Silva NI, Tennakoon DS, Karunarathna A, Gafforov Y, Pem D, Zhang SN, de Azevedo Santiago ALCM, Bezerra

JDP, Dima B, Acharya K, Alvarez-Manjarrez J, Bahkali AH, Bhatt VK, Brandrud TE, Bulgakov TS, Camporesi E, Cao T, Chen YX, Chen YY, Devadatha B, Elgorban AM, Fan LF, Du X, Gao L, Gonçalves CM, Gusmão LFP, Huanraluek N, Jadan M, Jayawardena RS, Khalid AN, Langer E, Lima DX, de Lima-Júnior NC, de Lira CRS, Liu JK, Liu S, Lumyong S, Luo ZL, Matočec N, Niranjana M, Oliveira-Filho JR, Papp V, Pérez-Pazos E, Phillips AJL, Qiu PL, Ren Y, Ruiz RFC, Semwal KC, Soop K, de Souza CAF, Souza-Motta CM, Sun LH, Xie ML, Yao YJ, Zhao Q, Zhou LW (2020) Fungal diversity notes 1277–1386: taxonomic and phylogenetic contributions to fungal taxa. *Fungal Diversity* 104 (1): 1-266. <https://doi.org/10.1007/s13225-020-00461-7>